Solid-State and Solution Structural Studies of 4-{[*C*(*E*)]-1*H*-Azol-1ylimino)methyl}pyridin-3-ols

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The new *N*-salicylideneheteroarenamines 1-4 were prepared by reacting the biologically relevant 3hydroxy-4-pyridinecarboxaldehyde (5) with 1*H*-imidazol-1-amine (6), 1*H*-pyrazol-1-amine (7), 1*H*-1,2,4triazol-1-amine (8), and 1*H*-1,3,4-triazol-1-amine (9). Solution ¹H-, ¹³C-, and ¹⁵N-NMR were used to establish that the hydroxyimino form **A** is the predominant tautomer. A combination of ¹³C- and ¹⁵N-CPMAS-NMR with X-ray crystallographic studies confirms that the same form is present in the solid state. The stabilities and H-bond geometries of the different forms, tautomers and rotamers, are discussed by using B3LYP/6-31G** calculations.

1. Introduction. – *N*-Salycilidenearenamines (salicylidene=(2-hydroxyphenyl)-methylene) show hydroxyimino **A**/oxoenamino **B** tautomerism, due to a proton-transfer process both in solution and in the solid state (*Scheme 1*) [1][2].



It has been established that in aromatic *Schiff* bases of *o*-hydroxybenzaldehydes, the equilibrium, normally favoring **A**, is shifted toward tautomer **B** when increasing the polarity of the solvent or the electron-withdrawing ability of the substituent R [1]. However, the N–H form **B** is rarely found [2][3].

Recently, we have been involved in the study of aromatic *Schiff* bases derived from 3-hydroxypyridine-4-carboxaldehyde (5) in an attempt to model the H-bonded structure of the cofactor pyridoxal-5'-phosphate (=3-hydroxy-2-methyl-5-[(phosphonooxy)methyl]pyridine-4-carboxaldehyde) intervening in various enzymatic transformations of amino acids [4]. As a continuation of that project, we decided to exploit

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our experience in the chemistry of *N*-aminoazoles [5-11] by preparing the *Schiff* bases **1–4** to study how the change in the nature of the substituent, heteroaromatic *vs.* aromatic, would affect the prototropic tautomerism of *Scheme 1*.

Moreover, the existence of additional H-bonding acceptors in the same molecule should modify the formation of the H-bonded network approaching what occurs in biological processes [12].

2. Results and Discussion. – The 4-[(1*H*-azol-1-ylimino)methyl] pyridin-3-ols 1-4 were prepared with good yields, by reaction of 3-hydroxypyridine-4-carboxaldehyde (5) with the corresponding 1*H*-azol-1-amines 6-9 [5] (*Scheme 2*).



Although (*E*)- and (*Z*)-isomers could be formed in the reaction, only the (*E*)-form was detected in all cases, as established by 2D-NOESY-NMR experiments showing the correlation between the imino proton and H–C(5) of the pyridine ring, as well as with H–C(5') of the azole moiety (or H–C(2') in the cases of 1 and 4) (*Sect. 2.2*). These results are in full agreement with B3LYP/6-31G** calculations that favor the (*E*)-isomer by 57.8 kJ mol⁻¹ in compound 1 (from 6), 46.7 kJ mol⁻¹ in 2 (from 7), 49.1 kJ mol⁻¹ in 3 (from 8) and 54.6 kJ mol⁻¹ in 4 (from 9). The same theoretical method was used to study the A/B tautomerism represented in *Scheme 1*: in the cases of compounds 1 and 4, the A tautomer with (*E*)-configuration is more stable than B by *ca.* 50 kJ mol⁻¹. The presence of an N-atom in the *a* position of the azolyl moiety, such as in compounds 2 and 3, further increases the difference in stability to the point that only tautomer A is a minimum. As we will show later on, these results are in agreement with the experimental observations.

The OH group of 1-4 can form an intramolecular H-bond with the N-atom, or intermolecular H-bonds with an N-atom of neighboring molecules. Moreover, there are 6 possible conformations for each 1-3 and 3 possible conformations for 4 due to rotations around the N–N, C–O, and C–C single bonds (*Fig. 1*). The results of the B3LYP/6-31G** calculations of the minimum energies of the various rotamers are summarized in *Table 1* showing that, in the case of isolated molecules in the gas phase, the **a** form presenting an intramolecular H-bond is always the most stable.

When looking at the azole moieties, only 2 and 3 have an N-atom at position N(2'), this being the reason why conformer **a** is favored. In compound **4**, conformers **a** and **b**, **c**



Fig. 1. Calculated conformations for 4-{[C(E)]-(1H-azol-1-ylimino)methyl}pyridin-3-ols 1-4 with atom numbering

and **d**, and **e** and **f** are pairwise identical, and finally in the imidazole derivative **1**, where all conformations are different, the relative stability order is $\mathbf{a} > \mathbf{b} > \mathbf{e} > \mathbf{f} > \mathbf{c} > \mathbf{d}$ (*Table 1*). In other words, an N-atom in the β -position of the azolyl moiety (series **1**) prefers the *s*-*trans* conformation by *ca*. 4.5 kJ mol⁻¹, in the α -position (series **2**) favors the *s*-*cis* conformation by *ca*. 24.5 kJ mol⁻¹, and in case of N-atoms in both the α - and β position (series **3**), the effects approximately add (*ca*. 26.0 kJ mol⁻¹).

2.1. Crystal and Molecular Structures. An X-ray study of all compounds was undertaken, but in the case of **2**, no appropriate crystals could be obtained. Crystals of **1**, **3**, and **4** consist in rotamers **e** interacting through intermolecular H-bonds, which led to different secondary structures. Selected bond distances and angles as well as the Hbond geometries are collected in *Table 2* and *3*, respectively. Helvetica Chimica Acta – Vol. 89 (2006)

		Α	b	c	d	e	f
1	+ZPE	-641.9652 (0.0) -641.7985 (0.0)	4.4 4.0	37.2 35.0	42.7 40.0	23.0 21.2	28.2 25.9
2	+ZPE	-641.9584 (0.0) -641.7914 (0.0)	23.01 21.84	38.5 36.4	68.0 64.2	26.8 25.0	51.2 48.1
3	+ZPE	- 658.0049 (0.0) - 657.8493 (0.0)	25.5 23.8	35.4 33.3	66.3 63.2	24.0 22.3	51.1 47.6
4	+ZPE	- 657.9832 (0.0) - 657.8289 (0.0)	0.0 0.0	34.2 32.5	34.2 32.5	20.3 19.1	20.3 19.1

Table 1. Absolute Energies [Hartree] and Relative Energies [kJ mol⁻¹]

	1	3	4		1	3	4
C(1)–O(1)	1.355(3)	1.340(2)	1.344(2)	O(1)–C(1)–C(5)	119.3(2)	119.1(2)	119.7(2)
C(1) - C(5)	1.402(3)	1.398(2)	1.389(3)	C(5)-C(1)-C(2)	118.4(3)	118.6(2)	118.8(2)
C(1) - C(2)	1.394(3)	1.393(3)	1.388(3)	C(1)-C(5)-C(6)	120.3(2)	118.7(2)	120.4(2)
C(2) - N(1)	1.336(3)	1.334(2)	1.331(3)	C(6)-N(2)-N(3)	117.3(2)	116.1(2)	116.8(2)
C(3) - N(1)	1.347(3)	1.340(2)	1.341(3)	C(1)-O(1)-H(1)	107.5	113.1	112.1
C(3) - C(4)	1.367(3)	1.374(3)	1.377(3)	O(1)-C(1)-C(2)-N(1)	179.6(3)	-179.0(2)	-179.4(2)
C(4) - C(5)	1.396(3)	1.393(3)	1.401(3)	O(1)-C(1)-C(5)-C(4)	178.6(2)	178.2(2)	179.9(2)
C(5) - C(6)	1.468(3)	1.465(3)	1.461(3)	C(5)-C(6)-N(2)-N(3)	179.9(2)	179.1(2)	177.6(2)
C(6) - N(2)	1.281(3)	1.276(2)	1.271(3)	N(4)-C(7)-N(3)-N(2)	-179.3(2)	-179.3(2)	178.2(2)
N(2) - N(3)	1.387(3)	1.386(2)	1.396(2)	C(6)-N(2)-N(3)-C(9)	2.9(5)	_	_
				C(6)-N(2)-N(3)-C(7)	-179.3(2)	-178.5(2)	174.0(2)
				C(6)-N(2)-N(3)-N(5)	-	2.9(3)	-
				C(6)-N(2)-N(3)-C(8)	-	-	-9.2(4)

Table 2. Selected Bond Lengths [Å] and Angles [°] for Compounds 1, 3, and 4

Table 3. Data of Hydrogen Bonds of Compounds 1, 3, and 4

	$D-H \cdots A$	δ (D–H) [Å]	$\delta(\mathbf{H}\cdots\mathbf{A})$ [Å]	$\delta(\mathbf{D}\cdots\mathbf{A})[\mathbf{\mathring{A}}]$	<(DHA) [°]
1	$O(1) - H(1) \cdots N(4)^{a}$	1.09	1.59	2.666(3)	166.0
3	$O(1) - H(1) \cdots N(1)^{b}$	0.95	1.69	2.616(2)	164.3
4	O(1) - H(1) - O(2)	1.08	1.55	2.615(2)	168.7
	$O(2) - H(2A) \cdots N(1)^{c}$	0.98	1.92	2.858(2)	158.2
	$O(2) - H(2B) \cdots N(5)^{d}$	1.00	1.79	2.761(2)	163.4

Molecular drawings of 1, 3, and 4 are presented in *Figs.* 2-4. All molecules are almost planar with the maximum deviation for 4 (*Table 2*), and the geometries are in agreement with those calculated for the corresponding conformers.

The H-bond in 1 and 3 lead to zig-zag chains in the [203] and [010] directions, respectively. In compound 4 the intermolecular H-bonds involve a H_2O molecule,

forming layers parallel to the (103) plane. In the three cases, the analysis of the intermolecular distances shows that the chains in **1** and **3** and the layers in **4** are independent.

2.2. *NMR Studies.* We recorded the ¹H-NMR spectra (*Fig. 1* for atom numberings) of all four compounds 1-4 in (D₆)DMSO solution, and analysis of the chemical shifts (see *Table 4*) led to the conclusion that only the hydroxyimino form **A** exists in all cases in the solvent (D₆)DMSO. Except for **2**, the 4-{[*C*(*E*)]-(1*H*-azol-1-ylimino)methyl}pyr-idin-3-ols were not soluble in CDCl₃.

In the case of 1*H*-imidazol-1-ylimino derivative **1**, we detected in the NOESY plot the correlation of the imino proton with H-C(5'), H-C(2'), and H-C(5) as well as that of OH with H-C(2) thus establishing the disruption of the intramolecular H-bond (*Fig.* 5).

Only in CDCl_3 , an intramolecular $\text{O}-\text{H}\cdots\text{N}=\text{C}$ H-bond was observed in the case of 1*H*-pyrazol-1-ylimino derivative **2** (*Fig* 6, *a*). In (D₆)DMSO, the NOESY plot (*Fig*. 6, *b*)



Fig. 2. Molecular and crystal structures of 1e



Fig. 3. Molecular and crystal structures of 3e



Fig. 4. Molecular and crystal structures of 4e

	Solvent	CH=N	H–C(2)	ОН	H–C(5)	H–C(6)
1	(D ₆)DMSO	8.99	8.36	10.70	7.66	8.15
	CD ₃ OD	9.05	8.28	a)	7.81	8.14
2	(D ₆)DMSO	9.38	8.35	10.73	7.71	8.14
	CDCl ₃	9.23	8.50	10.09	7.29	8.27
3	(D ₆)DMSO	9.38	8.38	10.92	7.72	8.16
4	(D ₆)DMSO	9.17	8.38	10.89	7.66	8.16
^a) Re	placed by OD.					

Table 4. ¹*H*-NMR Chemical Shifts of the Iminomethylpyridinol Moiety. δ in ppm.



Fig. 5. NOESY Plot ((D₆)DMSO) of 4-{[C(E)]-(1H-imidazol-1-ylimino)methyl}pyridin-3-ol (1)



Fig. 6. NOESY Plots of 4-[[C(E)]-(1H-pyrazol-1-ylimino)methyl]pyridin-3-ol (2): a) in $CDCl_3$ and b) in $(D_6)DMSO$

revealed a correlation between H–C(2) and OH, indicating the absence of intramolecular H-bonds in such a polar solvent [13]. For **2**, the major changes of the ¹H-NMR data (*Table 4*) in going from CDCl₃ to (D₆)DMSO affected H–C(5) (δ 7.29 to 7.71) and the OH (δ 10.09 to 10.73).

In the case of the 1H-1,2,4-triazol-1-ylimino derivative **3**, no correlations involving the imino proton could be detected, and in the case of the 1H-1,3,4-triazol-1-ylimino

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		CH=N	C(2)	C(3)	C(4)	C(5)	C(6)
1	(D ₆)DMSO	147.6 (J=170.5)	139.9 (J=179.1)	152.7	125.2	119.8 (J=163.8)	140.6 (J=181.1)
	CD ₃ OD	150.3	140.3	155.2	127.6	122.8	141.2
	CPMAS ^a)	144.4	138.1	154.4	124.6	119.2	141.1
2	(D ₆)DMSO	144.2 $(J = 172.2)$	139.9 (J=176.5)	152.9.	125.0	119.1 $(J = 160.1)$	140.4 $(J = 179.5)$
	CDCl ₃	150.3 (J = 172.1)	140.8 (J=179.9)	152.8	122.2	124.0 (J = 160.7)	141.2 $(J = 182.4)$
	CPMAS ^b)	140.7	138 ^h)	153.7	126.0	119.6	138 ^h)
		148.1		151.6	124.1	127.6	
	CPMAS ^c)	140.0	138 ^h)	154.3	129.0	120.3	138 ^h)
3	(D ₆)DMSO	147.8 (J=172.5)	140.1 (J=179.6)	153.1	124.2	119.1 $(J = 164.0)$	140.5 $(J = 182.6)$
	CPMAS ^d)	146.0	139.3	153.7	122.9	120.7	143.3
	CPMAS ^e)	147.4	143.0	155.9	124.0	119.9	143.0
			140.8	153.7	124.8		
					127.9		
4	(D ₆)DMSO	152.1 (J = 173.0)	140.1 (J=179.5)	152.9	124.5	119.5 (J = 166.7)	140.5 $(J = 182.3)$
	CPMAS ^f)	148.4	135.4	154.1	124.6	120.7	144.2
	CPMAS ^g)	152 ^h)	143 ^h)	155 ^h)	125 ^h)	121 ^h)	143 ^h)
		148 ^h)		152 ^h)	122 ^h)	119 ^h)	

Table 5. ¹³C-NMR Chemical Shifts and One-Bond Coupling Constants of the (Iminomethyl)pyridinol Moiety. δ in ppm, ¹J in Hz.

^{a)} Crystallized from toluene; δ 138.1 (C(2')), 124.6 (C(4')), 113.1 (C(5')). ^{b)} Crystallized from EtOH/H₂O; δ 137.6 and 135.4 (C(3')), 108.2 and 106.0 (C(4')), 132.6 and 130.6 (C(5')). ^{c)} Dissolved in CHCl₃ and evaporated; δ 134.0 (C(3')), 110.3 (C(4')), 129.3 (C(5')). ^{d)} Crystallized from toluene; δ 149.9 and 149.5 (C(3')), δ 138.0 (C(5')). ^{e)} Crystallized from H₂O; δ 151.4 and 147.4 (C(3')), δ 138.8 and 137.1 (C(5')). ^{f)} Crystallized from H₂O; δ 138.4 (C(2'), C(5')). ^{g)} Crystallized from CHCl₃/EtOH; δ 135.0 (C(2'), C(5')). ^h) Broad signal.

derivative 4, the NOESY experiments revealed a mixture of rotamers 4e (=4f)/4c (=4d).

Solution ¹³C-NMR studies (*Table 5*) showed signals for the C-atoms of the pyridine ring in agreement with the hydroxyimino structure (mean δ values: 150 (CH=N), 140 (C(2)), 153 (C(3)), 125 (C(4)), 119.5 (C(5)), and 140.5 (C(6)). The δ (C) for the azolyl substituents were within the normal ranges [5][7–9] (see *Exper. Part*). The CH ¹³C-NMR signals were found by gs-HMQC, and the quaternary C-atoms were assigned by long-range correlation experiments (gs-HMBC) [14]. In contrast to the *Schiff* bases we previously studied [4], the δ (C) of the C(6)s and their ¹J coupling constants (*Table 5*) were always larger than those of the C(2)s. For **2**, when changing from CDCl₃ to (D₆)DMSO, the most affected δ (C) (*Table 5*) were those of CH=N (δ 150.3 to 144.2) and of C(5) (δ 124.0 to 119.1), as expected for conformations **a** and **e**, respectively.

The ¹⁵N-NMR solution data were obtained by gs-HMBC (*Table 6*). The signals furnishing more information about the tautomerism in the 4-[(1*H*-azol-1-ylimino)methyl]-pyridin-3-ols **1**–**4** are those of the CH=N moiety, which appeared between $\delta(N)$ –64.8 and –75.8 in (D₆)DMSO, *i.e.*, at values typical for the nonprotonated N-atom of a *Schiff* base [1–4]. They confirmed that compounds **1**–**4** exist in the hydroxyimino tautomeric form **A**.

		CH=N	N(1)	N(1′)	N(2′)	N(3′)	N(4′)
1	(D ₆)DMSO	-66.9	- 52.9	-163.1		-120.8	
	CPMAS ^a)	-67.3	-56.0	-160.2		-142.5	
2	(D ₆)DMSO	-64.8	-53.1	-135.1	-94.5		
	CDCl ₃	-78.2	- 57.5	-141.7	-97.8		
	CPMAS ^b)	-78.9	-55.6	-134.3	-95.7		
			-63.8	-136.8			
	CPMAS ^c)	-85.7	-61.1	-133.5	-92.7		
3	(D ₆)DMSO	-72.4	-50.5	-129.4	-103.6		- 126.3
	CPMAS ^a)	-74.2	-47.9	-126.3	-101.7		- 138.9
	CPMAS ^d)	-84.0	-63.5	-127.6	-101.3		-138.3
		-79.3		-122.5			
		-75.2					
		-71.9					
4	(D ₆)DMSO	-75.8	-51.6	-164.9		-64.1	-64.1
	CPMAS ^e)	-86.3	-65.2	-162.5		-82.7	-82.7
	CPMAS ^d)	-75.7	55.2	-161.3		-71.7	-73.0
			59.7				

Table 6. ¹⁵N-NMR Chemical Shifts of 1-4. δ in ppm.

^a) Crystallized from toluene. ^b) Crystallized from EtOH/H₂O. ^c) Dissolved in CHCl₃ and evaporated. ^d) Crystallized from H₂O. ^e) Crystallized from CHCl₃/EtOH.

The main results of the ¹³C- and ¹⁵N-CPMAS-NMR studies in the solid state, will be discussed for each derivative. Thus compound **1**, an **e** conformer, presented a single signal for each nucleus in the ¹³C-CPMAS-NMR. As shown above, the X-ray structure of **1** confirmed the intermolecular associations involving the OH donor and the N(3') acceptor, thus affording an explanation for the increase of the ¹⁵N-NMR chemical shift $(\delta(\text{solid}) - \delta((D_6)\text{DMSO}) = -21.7$, see *Table 6*).

In the case of **2**, we did not succeed in growing suitable crystals for X-ray studies. Therefore, the solid-state NMR conclusions could not be confirmed by the X-ray structure. When **2** was dissolved in CHCl₃ and then the solution rapidly evaporated prior to the recording of the ¹³C-CPMAS-NMR spectrum (*Fig.* 7, *a*, NQS (non-quaternary suppression), and *Fig.* 7, *b*), only one structure was observed, most probably the one that presents the intramolecular H-bond, conformation **2a**. When **2** crystallized from an EtOH/H₂O mixture, two different rotamers **2a** and **2e** coexisted. This was clearly apparent from the C(3) and C(4) signals of the pyridine ring (*Fig.* 7, *c*, NQS) and C(4') of the pyrazole moiety (*Fig.* 7, *d*). The **2a** form evolved towards a mixture of **2a** and **2e** on standing.

The ¹³C-CPMAS-NMR spectrum of **3** obtained from toluene showed the presence of only structure **3e**, but in the spectrum registered after recrystallization from H_2O at least three distinct signals for C(4) were observed, attributable to the presence of other rotamers.

Crystals of **4** from H_2O corresponded to conformation **e**, but those obtained from CHCl₃/EtOH afforded two types of molecules, **4e** and **4c**, as clearly observed in the splitting of C(3) and C(4) in the ¹³C-CPMAS-NMR spectra.



Fig. 7. ¹³C-CPMAS-NMR Spectra of compound 2

3. Conclusions. – The structure of *Schiff* bases derived from 3-hydroxypyridine-4carboxaldehyde and four 1*H*-azol-1-amines was determined in the solid state (X-ray crystallography and CPMAS-NMR) and in solution (NMR and DFT calculations). Concerning tautomerism and E/Z isomerism about the C–N bond, all of them have the hydroxyimino structure with (*E*)-configuration. On the other hand, the conformation about the N–N and C–O single bonds strongly depends on the phase and the nature of the azolamine. In particular, in the solid state, O–H…N intermolecular Hbonds are always preferred to intramolecular ones.

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Experimental Part

General. M.p.: under a microscope Axiolab Zeiss with a TMS-92-Linkan heating stage and by DSC with a Seiko DSC 220C connected to a SSC5200H disk station; thermograms: sample size 0.003-0.010 g, scanning rate 2.0° min⁻¹. TLC: aluminium-backed plates of silica gel 60 F₂₅₄ (Merck, 0.2 mm). Elemental analyses: Perkin-Elmer-240 apparatus; performed by the 'Centro de Microanálisis Elemental-UCM', Madrid.

NMR Spectroscopy [14]. Solution NMR spectra: Bruker-DRX-400 (9.4 Tesla, 400.13 MHz for ¹H, 100.62 MHz for ¹³C and 40.56 MHz for ¹⁵N) spectrometer with a 5-mm inverse-detection H-X probe equipped with a z-gradient coil, at 300 K; chemical shifts δ in ppm rel. to the internal solvent CDCl₃ $(\delta(H) 7.26 \text{ and } \delta(C) 77.0), (D_6)DMSO (\delta(H) 2.49 \text{ and } \delta(C) 39.5), \text{ or } CD_3OD (\delta(H) 3.31 \text{ and } \delta(C) 39.5)$ 49.2) and rel. to the external standard nitromethane ($\delta(N)$ 0.00). Typical parameters for ¹H-NMR spectra: spectral width 4000 Hz, pulse width 7.5 μs , attenuation level 0 dB, resolution 0.15–0.25 Hz per point. Typical parameters for ¹³C-NMR spectra: spectral width 21 kHz, pulse width 10.6 µs, attenuation level -6 dB, resolution 0.6 Hz per point, relaxation delay 2 s; WALTZ-16 was used for broadband proton decoupling; the FIDs were multiplied by an exponential weighting (lb = 2 Hz) before Fourier transformation. 2D ¹H, ¹H-gs-COSY and inverse proton-detected heteronuclear shift-correlation spectra, ¹H, ¹³C-gs-HMQC, ¹H, ¹³C-gs-HMBC, and ¹H, ¹⁵N-gs-HMBC, were acquired and processed by using standard Bruker NMR software and in non-phase-sensitive mode. Gradient selection was achieved through a 5% sine truncated shaped pulse gradient of 1 ms. Selected parameters for ¹H, ¹H-gs-COSY: spectral width 2500-3500 Hz, acquisition data size 1024 points, one transient accumulated per increment, relaxation delay 1 s, a total of 256 experiments, data processing by using zero filling in the F1 domain and shifted sine-bell apodization of factor 0 in both dimensions. Selected parameters for ${}^{1}H$, ${}^{1}H$ -gs-NOESY: spectral width 2000-3000 Hz, acquisition data size 1024 points, 32 transients accumulated per increment, relaxation delay 1 s, mixing time 750-1800 ms, a total of 512 experiments, data processing by using zero filling in the F1 domain and shifted sine-bell apodization of factor 0 in both dimensions. Selected parameters for 1 H, 13 C-gs-HMQC and gs-HMBC spectra: spectral width 2500–3500 Hz for 1 H and 12.0–20.5 kHz for 13 C, 1024×256 data set, number of scans 2 (gs-HMQC), or 4 (gs-HMBC), and relaxation delay 1s. The FIDs were processed by using zero filling in the F1 domain and a sine-bell window function in both dimensions was applied prior to Fourier transformation. In the gs-HMQC experiments, GARP modulation of ¹³C was used for decoupling. Selected parameters for ¹H,¹⁵N-gs-HMBC spectra: spectral width 2500-3500 Hz for ¹H and 12.5 kHz for 15 N, 1024×256 data set, number of scans 4, relaxation delay 1s, 37–75 ms delay for the evolution of the ^{15}N ,¹H long-range coupling. The FIDs were processed by using zero filling in the F1 domain, and a sine-bell window function in both dimensions was applied prior to Fourier transformation.

Solid-state ¹³C- and ¹⁵N-CPMAS-NMR spectra: *Bruker-WB-400* spectrometer at 100.73 (¹³C) and 40.60 MHz (¹⁵N) and 300 K with a 4 mm *DVT* probehead. Samples were carefully packed in a 4-mmdiameter cylindrical zirconia rotor with Kel-F end-caps. Operating conditions involved 3.2 μ s 90° ¹H pulses and decoupling field strength of 78.1 kHz by TPPM sequence. The ¹³C spectra were originally referenced to a glycine sample and then the δ (C) were recalculated rel. to Me₄Si (carbonyl atom of glycine: δ 176.1). The ¹⁵N spectra were originally referenced to ¹⁵NH₄Cl and then converted to the nitromethane scale *via* the relationship δ (Me¹⁵NO₂)= δ (¹⁵NH₄Cl) – 338.1. Typical acquisition parameters for ¹³C-CPMAS-NMR: spectral width 40 kHz, recycle delay 60–120 s, acquisition time 30 ms, contact time 2–6 ms, and spin rate 12 kHz. To distinguish protonated and unprotonated C-atoms, the NQS (non-quaternary suppression) experiment by conventional cross-polarization was recorded; before the acquisition, the decoupler was switched off for a very short time of 25 μ s [15][16]. Typical acquisition parameters for ¹⁵N-CPMAS-NMR: spectral width 40 kHz, recycle delay 60–120 s, acquisition time 35 ms, contact time 8 ms, and spin rate 6 kHz.

DFT Calculations. The optimization of the structures of all compounds discussed in this paper was carried out at the hybrid B3LYP/6-31G** level [17][18] with basis sets of Gaussian-type functions by using Spartan '02 for Windows [19].

Syntheses. Compounds 1-4 were prepared by refluxing in toluene equimolar amounts of 5 [4] and the corresponding amine 6-9 [5] during 7 h and stirring overnight: yield 85-90%.

4-[[C(E)]-(1H-Imidazol-1-ylimino)methyl]pyridin-3-ol (1). TLC (CHCl₃/EtOH 9:1): R_f 0.31. The crystals were purified by crystallization (C₇H₈). M.p. 251.4° (dec; DSC); under the microscope, **1** changed its appearance at 219° decomposing at 263°. ¹H-NMR ((D₆)DMSO): 10.70 (br. *s*, OH); 8.99 (*s*, CH=N); 8.36 (*s*, H-C(2)); 8.17 (*t*, ⁴J(2',4') = ⁴J(2',5') = 1.3, H-C(2')); 8.15 (*d*, ³J(5,6) = 5.0, H-C(6)); 8.04 (*t*, ³J(4', 5') = ⁴J(2',5') = 1.3, H-C(5')); 7.66 (*d*, H-C(5)); 7.07 (*t*, H-C(4')). ¹³C-NMR ((D₆)DMSO): 152.7 (³J=³J=4.7, C(3)); 147.6 (¹J=170.5, ³J=4.5, CH=N); 140.6 (¹J=181.1, ³J=10.9, C(6)); 139.9 (¹J=179.1, ³J=11.2, C(2)); 136.4 (¹J=213.6, C(2')); 128.8 (¹J=190.7, ³J=11.8, ²J=9.3, C(4')); 125.2 (C(4)); 119.8 (¹J=163.8, ³J=9.5, ²J=3.5, C(5)); 112.8 (¹J=195.6, ³J=2.7, ²J=16.9, C(5')). ¹H-NMR

(CD₃OD): 9.05 (*s*, CH=N); 8.28 (*s*, H–C(2)); 8.18 (*t*, ${}^{4}J(2',4') = {}^{4}J(2',5') = 1.4$, H–C(2')); 8.14 (*d*, ${}^{3}J(5, 6) = 5.1$, H–C(6)); 7.92 (*t*, ${}^{3}J(4',5') = 1.4$, H–C(5')); 7.81 (*d*, H–C(5)); 7.13 (*t*, H–C(4')). 13 C-NMR (CD₃OD): 155.2 (C(3)); 150.3 (CH=N); 141.2 (C(6)); 140.3 (C(2)); 138.0 (C(2')); 129.6 (C(4')); 127.6 (C(4)); 122.8 (C(5)); 113.6 (C(5')). Anal. calc. for C₉H₈N₄O: C 57.44, H 4.28, N 29.77; found: C 57.35, H 4.34, N 29.84.

4-[[C(E)]-(1H-Pyrazol-1-ylimino)methyl]pyridin-3-ol (2). TLC (CHCl₃/EtOH 9:1): R_f 0.79. The crystals were purified by crystallization (CHCl₃). M.p. 163° (microscope) and 164.4° (DSC). ¹H-NMR ((D₆)DMSO): 10.73 (br. *s*, OH); 9.38 (*q*, ⁴*J*(CH,5)=0.7, ⁵*J*(CH,6)=0.7, ⁶*J*(CH,3')=0.7, CH=N); 8.35 (*d*, ⁴*J*(2,6)=0.4, ⁵*J*(2,5)=0.6, H-C(2)); 8.14 (*d*dd, ³*J*(5,6)=5.0, H-C(6)); 8.08 (*d*d, ³*J*(4',5')=2.5, ⁴*J*(3',5')=0.8 H-C(5')); 7.71 (*t*, H-C(5)); 7.68 (*t*d, ³*J*(4',3')=1.9, H-C(3')); 6.51 (*d*d, H-C(4')). ¹³C-NMR ((D₆)DMSO): 152.9 (C(3)); 144.2 (¹*J*=172.2, CH=N); 140.4 (¹*J*=179.5, C(6)); 139.9 (¹*J*=176.5, C(2)); 138.5 (¹*J*=187.8, C(3')); 129.9 (¹*J*=195.0, C(5')); 125.0 (C(4)); 119.1 (¹*J*=160.1, C(5)); 107.0 (¹*J*=178.7, C(4')). ¹H-NMR (CDCl₃): 10.09 (br. *s*, OH); 9.23 (*d*, ⁶*J*(CH,3')=0.8, CH=N); 8.50 (*s*, H-C(2)); 8.27 (*d*, ³*J*(5,6)=4.9, H-C(5)); 6.45 (*d*d, H-C(4')). ¹³C-NMR (CDCl₃): 152.8 (C(3)); 150.3 (¹*J*=172.1, ³*J*=6.4, CH=N); 141.2 (¹*J*=182.4, ³*J*=10.1, C(6)); 140.8 (¹*J*=179.9, ³*J*=10.4, C(2)); 139.0 (¹*J*=188.3, ³*J*=9.1, ²*J*=5.4, C(3')); 129.0 (¹*J*=192.4, ³*J*=3.7, ²*J*=8.8, C(5')); 124.0 (¹*J*=160.7, ³*J*=9.1, C(5)); 122.2 (C(4)); 107.5 (¹*J*=179.8, ²*J*=²*J*=8.8, C(4')). Anal. calc. for C₉H₈N₄O: C 57.44, H 4.28, N 29.77; found: C 57.30, H 4.68, N 27.71.

	1	3	4
Identification code	CCDC-285402	CCDC-285403	CCDC-285404
Empirical formula	$C_9H_8N_4O$	$C_8H_7N_5O$	$C_8H_9N_5O_2$
Formula weight	188.19	189.19	207.20
Crystal system	Monoclinic	Monoclinic	Monoclinic
Space group	P2(1)/n	P2(1)/c	P2(1)/n
Unit cell dimensions			
<i>a</i> [Å]	5.2731(8)	4.6828(6)	4.8932(5)
<i>b</i> [Å]	14.389(2)	8.488(1)	22.033(2)
c [Å]	11.677(2)	21.990(3)	9.169(1)
β [°]	93.735(4)	90.308(3)	97.697(2)
Volume [Å ³]	884.1(2)	874.0(2)	979.6(2)
Ζ	4	4	4
Density (calculated) [Mg/m ³]	1.414	1.438	1.405
Absorption coefficient [mm ⁻¹]	0.099	0.104	0.106
F(000)	392	392	432
Scan Technique	ω and φ	ω and φ	ω and φ
θ range for data collection [°]	2.25 to 24.99	1.85 to 24.99	1.85 to 24.99
Index ranges	$-6 \le h \le 6$	$-5 \le h \le 5$	$-5 \le h \le 5$
	$-17 \leq k \leq 16$	$-10 \le k \le 10$	$-23 \le k \le 26$
	$-7 \leq l \leq 13$	$-17 \le l \le 26$	$-8 \le l \le 10$
Reflections collected	4549	4418	5031
Independent reflections	1553 (R(int) = 0.072)	1541 (R(int) = 0.0442)	1716 (R(int) = 0.0462)
Data, restraints, parameters	1553, 0, 136	1541, 0, 134	1716, 0, 145
Reflections observed $(I > 2\sigma(I))$	749	1011	1014
Goodness-of-fit on F^2	0.913	0.931	0.923
<i>R</i> ^a) [observed reflec.]	$R_1 = 0.0447$	$R_1 = 0.0396$	$R_1 = 0.0421$
$Rw_{\rm F}^{\rm b}$) (all data)	$wR_2 = 0.1078$	$wR_2 = 0.1018$	$wR_2 = 0.1089$
^{a)} $\Sigma F_{o} - F_{c} / \Sigma F_{o} $. ^b) { $\Sigma [w(F_{o}^{2} - V_{o}^{2})]$	$-F_{\rm c}^2)^2]/\Sigma[w(F_{\rm o}^2)^2]\}^{1/2}.$		

4-[[C(E)]-(1H-1,2,4-Triazol-1-ylimino)methyl]pyridin-3-ol (**3**). TLC (CHCl₃/EtOH 9:1): R_f 0.55. The crystals were purified by crystallization (CHCl₃/EtOH). M.p. 230° (microscope), and 236.6° with decomposition at 263.6° (DSC). ¹H-NMR ((D₆)DMSO): 10.92 (br. *s*, OH); 9.38 (*s*, CH=N); 8.99 (*s*, H-C(5')); 8.38 (*s*, H-C(2)); 8.20 (*s*, H-C(3')); 8.16 (*d*, ³*J*(5,6)=5.0, H-C(6)); 7.72 (*d*, H-C(5)). ¹³C-NMR ((D₆)DMSO): 153.1 (C(3)); 150.0 (¹*J*=211.1, ³*J*=12.4, C(3')); 147.8 (¹*J*=172.5, ³*J*=3.6, CH=N); 140.5 (¹*J*=182.6, ³*J*=10.9, C(6)); 140.1 (¹*J*=179.6, ³*J*=11.2, C(2)); 134.1 (¹*J*=218.7, ³*J*=6.5, C(5')); 124.2 (³*J*=³*J*=²*J*=6.3, C(4)); 119.1 (¹*J*=164.0, ³*J*=9.9, ²*J*=4.2, C(5)). Anal. calc. for C₈H₇N₅O: C 50.79, H 3.73, N 37.02; found: C 50.07, H 4.01, N 35.35.

4-[[C(E)]-(1H-1,3,4-Triazol-1-ylimino)methyl]pyridin-3-ol (4). TLC (CHCl₃/EtOH 9:1): R_f 0.11. The crystals were purified by crystallization (H₂O/EtOH). M.p. 280° (microscope) and 240° (dec.; DSC). ¹H-NMR ((D₆)DMSO): 10.89 (br. *s*, OH); 9.23 (*s*, H–C(2'), H–C(5')); 9.17 (*s*, CH=N); 8.38 (*s*, H–C(2)); 8.16 (*d*, ³*J*(5,6)=5.0, H–C(6)); 7.66 (*d*, H–C(5)). ¹³C-NMR ((D₆)DMSO): 152.9 (C(3)); 152.1 (¹*J*=173.0, CH=N); 140.5 (¹*J*=182.3, ³*J*=11.4, C(6)); 140.1 (¹*J*=179.5, ³*J*=11.3, C(2)); 139.1 (¹*J*=216.1, ³*J*=3.3, C(2'), C(5')); 124.5 (C(4)); 119.5 (¹*J*=166.7, C(5)). Anal. calc. for C₈H₇N₅O.H₂O: C 46.38, H 4.38, N 33.80; found: C 46.47, H 4.41, N 34.00.

X-Ray Data Collection and Structure Refinement. Suitable crystals for X-ray diffraction experiments were obtained by crystallization from H₂O/EtOH. Data collection for compounds were carried out at r.t. with a *Bruker-Smart-CCD* diffractometer by using graphite-monochromated Mo- K_a radiation (λ 0.71073 Å) operating at 50 kV and 30 mA. In all cases, data were collected over a hemisphere of the reciprocal space by combination of three exposure sets. Each exposure of 30s covered 0.3 in ω . The cell parameter were determined and refined by a least-squares fit of all reflections. A summary of the fundamental crystal and refinement data of 1, 3, and 4 is given in *Table 7*. The structures were solved by direct methods (SHELXS-97) and refined by full-matrix least-square procedures on F^2 (SHELXL-97) [20]. All non-H-atoms were refined anisotropically. All H-atoms were located on a difference *Fourier* map and refined riding on the respective C- or O-atoms. Largest peaks and holes in the final difference map were 0.165 and -0.150, 0.175 and -0.178, 0.154 and -0.151 e Å⁻³ for 1, 3, and 4, respectively.

CCDC-285402, CCDC-285403, and CCDC-285404 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge *via* http://www.ccdc.cam.ac.uk/data_request/cif from the *Cambridge Crystallographic Data Centre*.

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